

Virulence Factors and Pathogenicity development of *Shigella* , Review

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Abstract

Shigella is a main cause of gastroenteritis and it is responsible for 5 to 10 % of diarrheathrough the world. The natural hosts of *Shigella* are typically humans and other primates, but it has been shown that the host range of *Shigella* has expanded to many animals. Although *Shigella* is becoming a major threat to animals, there is limited information on the genetic background of local strains. *Shigella* is the major cause of bacillary dysentery world-wide. It is divided into four species, named *S. flexneri*, *S. sonnei*, *S. dysenteriae*, and *S. boydii*, which are distinct genomically and in their ability to cause disease. Shigellosis, the clinical presentation of *Shigella* infection, is characterized by watery diarrhea, abdominal cramps, and fever. *Shigella*'s ability to cause disease has been attributed to virulence factors, which are encoded on chromosomal pathogenicity islands and the virulence plasmid. However, information on these virulence factors is not often brought together to create a detailed picture of infection, and how this translates into shigellosis symptoms. Firstly, *Shigella* secretes virulence factors that induce severe inflammation and mediate enterotoxin effects on the colon, producing the classic watery diarrhea seen early in infection. Secondly, *Shigella* injects virulence effectors into epithelial cells via its Type III Secretion System to subvert the host cell structure and function. This allows invasion of epithelial cells, establishing a replicative niche, and causes erratic destruction of the colonic epithelium. Thirdly, *Shigella* produces effectors to down-regulate inflammation and the innate immune response.

Key words : Virulence Factors , Antibiotic Resistance , *Shigella* , Pathogenicity .distinct genomically

Introduction

Diarrheal diseases caused by bacterial, viral, or parasitic pathogens are a major public health problem. Estimations by the World Health Organization (WHO) indicate that the world population suffered from 4.5 billion incidences of diarrhea causing 1.8 million deaths in the year 2002 (334). Approximately 99% of the cases occurred in developing countries, where poor hygiene and limited access to clean drinking water promote the spread of enteric diseases. Malnutrition and the lack of appropriate medical intervention contribute to the high mortality rate, especially for young children. Species of the genus *Shigella* are among the bacterial pathogens most frequently isolated from patients with diarrhea. Five to fifteen percent of all diarrheal episodes worldwide can be attributed to an infection with *Shigella*, including 1.1 million fatal cases (136). Two-thirds of all episodes and deaths occur in children under 5 years.

Shigellosis or blood dysentery is widespread in underdeveloped or developing regions with poor hygiene and limited access to clean drinking water and has become a serious threat to public health . Shigellosis is caused by non-motile, facultative anaerobic gram-negative bacilli of the Enterobacteriaceae family, including *S. dysenteriae*, *S. flexneri*, *S. boydii*, and *S. sonnei*. *Shigella* species have high effectiveness in invasive systems that enable bacteria to invade and multiply within the human intestinal epithelia, ultimately leading to severe inflammatory colitis, which is referred to as bacillary dysentery or shigellosis . Various virulence factors located on chromosomes or large virulence plasmids are recognized as crucial factors related to the pathogenesis of shigellosis. Moreover, these different virulence factors are associated with the colonization of intestinal cells and intracellular invasion, which may partly explain why various manifestations are detected in the clinic, such as intestinal inflammatory responses and watery diarrhea. Bacterial cell-to-cell movement and dissemination within epithelial cells of the intestine are allowed by the *ipH* gene, which is encoded by chromosomal DNA and/or recombinant plasmids, while *ial*, which is encoded by plasmids (invasion-associated loci), enables *Shigella* bacteria to penetrate intestinal epithelial tissues . The chromosomal genes *set1A* and *set1B* encode *Shigella* enterotoxin 1 (ShET-1) , which is easily detected in all *S. flexneri* 2a isolates.

OspB: Promotes PMN Migration, Inflammation, and Cell Proliferation

Like most effectors, OspB is found in the four *Shigella* species, and has homolog in *Salmonella* species. Although its biochemical function is unknown, it is thought that OspB plays a role in the activation of extracellular-signal-regulated kinases (ERK) and p38 MAPK pathways, resulting in phosphorylation of phospholipase A2 and the generation of eicosanoids. OspB is capable of nuclear localization for activation of MAPK signaling pathways. This contributes to inflammation and PMN migration, possibly inducing hepxilin A3, an arachidonic acid derivative, and apical secretion of IL-8, a PMN chemoattractant. An ospB⁻ mutant had a 60% decrease in PMN migration and a 30% decrease in ERK1/2 activation 90 min post-infection when compared to wild-type *Shigella*. Furthermore, it showed that an ospB⁻ knockout displayed significantly reduced onset and severity of symptoms in the guinea pig keratoconjunctivitis model of infection (Sereny test). However, OspB also activates the master regulator of cell growth mTOR via a direct interaction with the cellular scaffold protein IQGAP1, which also interacts with mTOR activators ERK1/2. This seems to restrict the spread of *S. flexneri* in cell monolayers, possibly by enhancing cell proliferation in infected foci.

OspC1: Promotes PMN Migration and Inflammation

OspC1 is part of the ospC family. There is 96% identity between ospC2, ospC3, and ospC4, but only 74% identity between these three ospC genes and ospC1. This level of similarity may indicate redundancy. However, ospC4 is a pseudogene and different functions have been identified for OspC1 and OspC3 (discussed later). Tagged OspC1 is found throughout the host cytoplasm, localizing primarily to the nucleus. An ospC1⁻ knockout showed a significant decrease in the amount of neutrophil recruitment to the epithelial cells in PMN migration assays, which was restored to wild-type levels on complementation with a plasmid expressing ospC1. It showed that this increase in PMN migration correlated with increase in the phosphorylation of ERK1/2 pathways mediated by OspC1. An ospC1⁻ knockout showed a decrease in phosphorylation of ERK1/2 compared to wild-type levels but no reduction in IL-8 secretion. OspC1 plays a role in *Shigella* virulence in vivo as an ospC1⁻ knockout had reduced amounts of swelling and inflammation in the Sereny test, with clearance of infection after 2 days.

OspZ: Promotes PMN Migration and Inflammation

In *S. flexneri* 2a, an ospZ⁻ knockout has no effect on the Sereny test. However, an ospZ⁻ knockout caused a significant decrease in PMN migration. The knockout also had 63 and 53% ERK1/2 phosphorylation and NFκB activation, respectively, when compared to wild-type *S. flexneri*. OspZ therefore plays a role in the migration of PMN leukocytes across the epithelial barrier. However, it was discovered that *S. flexneri* species, excluding *S. flexneri* serotype 6, contain a stop codon at amino acid 188, forming a truncated protein lacking an IDSYMK motif at position 209. The full length OspZ proteins in the remaining *Shigella* species were found to have an immunosuppressive function through prevention of NFκB activation. Finally, an OspZ homolog, NleE, is found in enteropathogenic *Escherichia coli* (EPEC), and both NleE and OspZ can substitute for each other.

Serine Protease Autotransporters of Enterobacteriaceae

Serine Protease Autotransporters of Enterobacteriaceae (SPATEs) are a family of proteases which catalyze their own secretion via the Type V secretion pathway. *Shigella* has three known SPATEs, not all of which are found in each species. Their secretion is thermoregulated (37°C) and pH-dependent. They have different proposed activities relevant to intestinal penetration: induction of mucin secretion and cleavage (Pic), destabilization of focal adhesions via cleavage of fodrin (SigA), and, through unknown targets, enterotoxicity, fluid accumulation and epithelial desquamation (SigA and SepA).

Shigella Enterotoxin 1 and Shigella Enterotoxin 2:

Enterotoxin Activity in the Jejunum Shigella enterotoxin 1 (ShET1) and Shigella enterotoxin 2 (ShET2) are virulence determinants proposed to mediate early fluid secretion in the jejunum to establish infection in the colon and produce the characteristic watery diarrhea seen early in shigellosis. The shared name is due to their similar properties as enterotoxins, as there is no homology between ShET1 and ShET2. ShET1 is encoded by set1A and set1B genes on the Shigella chromosome as part of the SHI-1 PAI, and only present in *S. flexneri* 2a isolates. The two subunits are proposed to form a holotoxin complex in an A1-B5 configuration, producing a 55 kDa complex. The holotoxin may follow a secretion mechanism similar to that of the cholera holotoxin, via the Sec pathway and Type II secretion. When ion transport across a cultured epithelium was measured in an Ussing chamber, a set1AB- knockout had 60% lower Isc (shortcircuit current) in comparison to wild-type strains. The effect on Isc of ShET1 was also dose-dependent, and washout of ShET1 produced no change in Isc, indicating irreversible binding of ShET1 to epithelial receptors. However, the set1A and set1B genes overlap with the pic gene but are divergently transcribed. Therefore, the additional pic- knockout may have caused these effects. Complementation of the pic/set1AB mutant with pic and set1AB individually, showing that pic has a more significant contribution to restoring Isc levels to wild-type, although set1AB complementation also produced a significant increase in Isc. Figure 1 and table 1 showed role of virulence factors and genes.

Adhesion to the Colonic Epithelium at the Basolateral Surface
Lipopolysaccharide: Glucosylation for T3SS Accessibility

The lipopolysaccharide (LPS) is a common feature of Gram-negative pathogens, triggering the host immune response and inflammatory reactions during infection. LPS modification by glucosylation is thought to contribute to Shigella adhesion and invasion by revealing the T3SS for efficient activation upon contact with the host cell. Studies showed that glycosyltransferase gtrA- and gtrB- mutants had only a partial conversion of the O-antigen serotype, and a gtrX- mutant had no conversion at all. A mutation in the gtr operon leads to a reduced ability to invade, and this invasion is restored when the gtr operon is reintroduced. The reduction in O-antigen length by glucosylation enhances accessibility of the T3SS for contact with the host epithelial cell to initiate invasion. IpaB: Binds CD44 at the Basolateral Surface, IpaC: Binds CD44 at the Basolateral Surface, IpaD: Binds CD44 at the Basolateral Surface, IpgC: Binds CD44 at the Basolateral Surface, OpsE1/E2: Bile Salts-Dependent Adhesion, IcsA (VirG): Polar Adhesion.

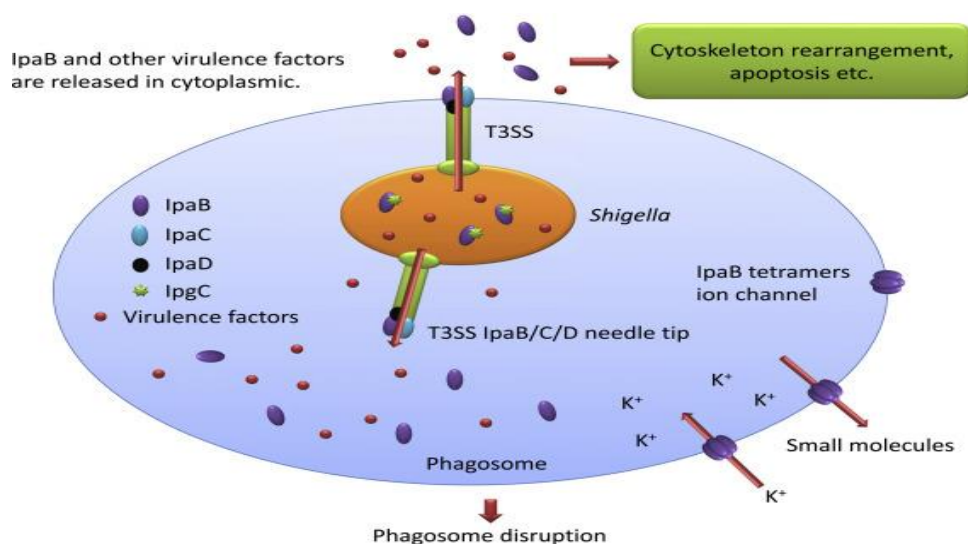


Figure-1: Shown role virulence factors in *Shigella*

Table-1: Shown genes of virulence factors in *Shigella*

PAI	Gene(s)	Protein function	References
SHI-1	<i>sigA</i>	Putative enterotoxin	Al-Hasani et al., 2009
	<i>pic</i>	Intestinal colonization	Navarro-Garcia et al., 2010
	<i>set 1A, set 1B</i>	ShET1 enterotoxin	Fasano et al., 1997
SHI-2	<i>iucA-D</i>	Siderophore, complexes with iron	Vokes et al., 1999
	<i>iutA</i>	Bacterial receptor for iron-siderophore complex	Vokes et al., 1999
	<i>shiA-G</i>	Novel ORFS, ShiA involved in reduction in host inflammatory response	Ingersoll et al., 2003
SHI-3	<i>iucA-D</i>	Siderophore, complexes with iron	Purdy and Payne, 2001
	<i>iutA</i>	Bacterial receptor for iron-siderophore complex	Purdy and Payne, 2001
SHI-O	<i>gtrA, gtrB, gtr</i>	Serotype conversion and O-antigen modification	Allison and Verma, 2000
Stx-phage P27	<i>stxAB</i>	Shiga toxin	Yang et al., 2005

Table 3- Genes location of virulence factors in *Shigella*

SPATE	Gene location	Putative function	Role in infection	References
Pic	SHI-1 (opposite <i>set1AB</i>)	Cleavage of mucin	Penetrate colonic mucus layer to access epithelium	Gutierrez-Jimenez et al., 2008
		Mucin secretagogue	Mucus-containing dysentery in shigellosis	Navarro-Garcia et al., 2010
SigA	SHI-1	Cytopathic activity	Cleavage of fodrin to destabilize links between actin cytoskeleton and membrane proteins, detachment of focal adhesions	Canizalez-Roman and Navarro-García, 2003; Al-Hasani et al., 2009
		Enterotoxigenic activity	Fluid accumulation	Al-Hasani et al., 2000
SepA	Virulence plasmid	Enterotoxigenic activity	Fluid accumulation	Benjeloun-Touimi et al., 1995
		Epithelial desquamation	Disease progression	Coron et al., 2009

Lysis of the Double Membrane Vacuole Vps and VacJ: Proposed ABC Transporter

The *vpsABC* operon is found on the *Shigella* chromosome, and consists of VpsA, a possible ATP-Binding Cassette(ABC) transporter protein, and VspB and VspC, proposed transmembrane proteins. Both *vpsC*- and *vspA*- knockout had a defect in plaque formation but were similar to wild type strains in their capability to invade, indicating that they play a role in intercellular spread. VacJ is also encoded on the chromosome, and a *vacJ*- knockout is incapable of escaping into the recipient cell cytoplasm, suggesting that VacJ also plays a role in intercellular spread. Describe a Vps/VacJ ABC transporter, which maintains asymmetry of lipids in the outer membrane and in the context of *Shigella* infection is required for lysis of the double membrane vacuole. Transformation of *vps/vacJ* knockouts with a plasmid expressing *pIdA*, a phospholipase in other Gram negative bacteria, was able to restore the maintenance of outer membrane lipid asymmetry but was unable to lyse the double membrane vacuole, indicating that these two functions of the proposed Vps/VacJ ABC transporter are separate. Another substrate may therefore be transported across the membrane to induce vacuolar lysis, however this is yet to be discovered.

Virulence Determinants in Disease-Causing Species

S. flexneri has a relatively stable genome, and acquired the virulence plasmid early in its evolution. *S. flexneri* is capable of persisting in water for several months, similarly to *Vibrio cholera*. This may explain its epidemiological prevalence, as access to human hosts for months at a time could facilitate the endemics seen in countries with poor water sanitation. Speculatively, it may cause the most disease as *S. flexneri* 2a harbors the SHI-1 PAI, which encodes Pic, SigA, and ShET1. Pic may confer an advantage for scavenging nutrients, therefore other strains may have a metabolic disadvantage when compared to *S. flexneri*. The importance of Pic and ShET1 in pathogenesis has also been highlighted by . The deletion of *pic* and *set1AB*, in addition to *ospD3*, in a guanine autotrophic (*guaAB*-) background produced an increasingly attenuated vaccine, with none of the 14 volunteers developing diarrhea. The truncated OspZ found in *S. flexneri* has a pro-inflammatory role, compared to its anti-inflammatory in the remaining sub-species, which may confer an inflammatory advantage for initial establishment of infection .

Conclusion

Gene expression of virulence genes in *Shigella* play important role in spreading pathogenicity like *sigA*, *invE* and *virF* genes showed that this classical regulatory pathway of *Shigella* virulence factor gene expression can play a major role in the pathogenesis of this bacterium. This bacterium consider originating from harmless enterobacterial relatives, the structure and function of the major virulence factors, including a T3SS and cognate effector proteins, as well as the interactions of *S. flexneri* with various host cells. All these mechanisms of virulence factors production under control quorum sensing signals.

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